

# Rapid multi-locus tNGS-based genotyping of *Listeria monocytogenes* by the Clear Safety™ automated platform to identify outbreak-relevant isolates

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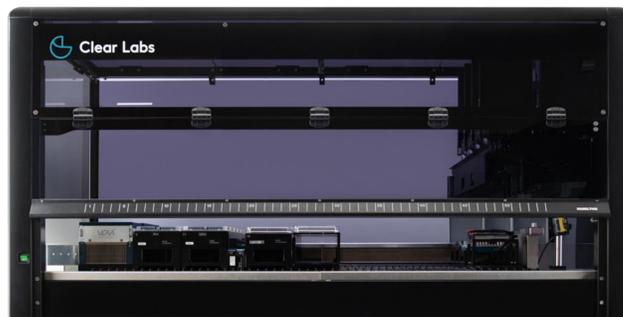
## INTRODUCTION

*Listeria monocytogenes* is a prevalent food-borne pathogen and a leading cause of recalls in the food industry due to its severe public health risks and invasiveness in the production environment. Quickly tracking subtype patterns in *Listeria monocytogenes* is becoming increasingly necessary to get ahead of contamination events and potential outbreaks.

Current *Listeria* strain typing methods include Whole Genome Sequencing (WGS), Pulsed-field gel electrophoresis (PFGE), and Multiple loci Variable-number tandem repeat analysis (MLVA), and require colony isolation, experienced lab personnel, and several days to provide results.

The Clear Safety *Listeria* platform is an automated targeted Next Generation Sequencing (tNGS) platform which can amplify and sequence multiple loci in the *Listeria* genome to rapidly and easily speciate and strain type *Listeria* in isolates and real-world samples.

To assess the accuracy and ease of the Clear Safety *Listeria* product, *L. monocytogenes* strains associated with 3 distinct and impactful food-borne outbreaks were run on the platform amongst other samples representing similar but distinct subtypes, and different as controls.



## METHODS

### Culture Incubation and Preparation

Sixteen *Listeria monocytogenes* strains isolated from clinical and food sources related to three different outbreaks (Nova Scotia, 1981; Massachusetts, 1983; and California, 1985) and 9 ATCC control *Listeria spp.* strains were included in this study. All strains were sampled directly from glycerol stocks, grown in 2mL of Clear *Listeria* Media and incubated at 35°C overnight.

50µL of each overnight culture was added to 450µL of Buffer in duplicate and loaded onto the end-to-end automated system which performs cell lysis, PCR amplification of multiple *Listeria* loci, library preparation, and sequencing on an Oxford Nanopore Technologies (ONT) flow cell. Amplicon sequences were demultiplexed, quality-filtered, and mapped to a reference database through an automated workflow to confirm the species and strain type of the *Listeria* isolates.

### Statistical Methods

Concordance between genotyping results and reference metadata provided at strain acquisition was evaluated using Cohen's Kappa statistic.

The Hunter-Gaston Discriminatory Index (HGDI) was used to quantify the probability that two randomly selected strains would be distinguishable based on their genotypes.

*L. grayi* was excluded from the statistical analyses as a *Listeria* species which is inherently difficult to type beyond the species level.

Table 1. Sample Concordance Counts Between Outbreak Collection Data and Clear Safety *Listeria* Results

Outbreak Metadata	Clear Safety Results					Total
	Halifax 1981	Mass. 1983	Calif. 1985	Other		
Halifax 1981	10	0	0	0	10	
Mass. 1983	0	8	0	0	8	
Calif. 1985	0	0	12	0	12	
Other	2	2	0	14	18	
<b>Total</b>	<b>12</b>	<b>10</b>	<b>12</b>	<b>14</b>	<b>48</b>	

Figure 1: *Listeria spp.* Tested with Metadata and Clear Safety Results

Random sample #	Genus	species	Source	Collection	Serotype	Clear Safety <i>Listeria</i>	
						Rep 1	Rep 2
1	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate California 1985	USDA-ARS	4b	A	A
2	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate Throat Halifax, Nova Scotia 1981	USDA-ARS	4b	B	B
3	<i>Listeria</i>	<i>monocytogenes</i>	Poultry, England	Microbiologics	1/2a	D	D
4	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate, Massachusetts, 1983	USDA-ARS	4b	C	C
5	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate, Massachusetts, 1983	USDA-ARS	4b	C	C
6	<i>Listeria</i>	<i>grayi</i>	Standing corn stalks and leaves	Microbiologics	NA	N/A	N/A
7	<i>Listeria</i>	<i>seeligeri</i>	Soil	Microbiologics	NA	E	E
8	<i>Listeria</i>	<i>monocytogenes</i>	Spinal fluid from child with meningitis, Germany	Microbiologics	4b	C	C
9	<i>Listeria</i>	<i>monocytogenes</i>	Poultry, England	Microbiologics	4e	B	B
10	<i>Listeria</i>	<i>monocytogenes</i>	Jalisco Cheese, California 1985	USDA-ARS	4b	A	A
11	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate California 1985	USDA-ARS	4b	A	A
12	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate Stillborn, Halifax, Nova Scotia 1981	USDA-ARS	4b	B	B
13	<i>Listeria</i>	<i>monocytogenes</i>	Cole Slaw Halifax, Nova Scotia 1981	USDA-ARS	4b	B	B
14	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate, Massachusetts, 1983	USDA-ARS	4b	C	C
15	<i>Listeria</i>	<i>monocytogenes</i>	Raw Milk, Massachusetts, 1983	USDA-ARS	4b	F	F
16	<i>Listeria</i>	<i>welshimeri</i>	Decaying plant material	Microbiologics	6b	G	G
17	<i>Listeria</i>	<i>monocytogenes</i>	Human spinal fluid, Scotland	Microbiologics	2	H	H
18	<i>Listeria</i>	<i>monocytogenes</i>	Halifax, Nova Scotia 1981	USDA-ARS	4b	B	B
19	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate Vagina Halifax, Nova Scotia 1981	USDA-ARS	4b	B	B
20	<i>Listeria</i>	<i>monocytogenes</i>	Animal tissue, 1931	Microbiologics	4a	I	I
21	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate California 1985	USDA-ARS	4b	A	A
22	<i>Listeria</i>	<i>innocua</i>	Cow brain	Microbiologics	6a	J	J
23	<i>Listeria</i>	<i>monocytogenes</i>	Jalisco Cheese, California 1985	USDA-ARS	4b	A	A
24	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate, Massachusetts, 1983	USDA-ARS	4b	C	C
25	<i>Listeria</i>	<i>monocytogenes</i>	Clinical Isolate California 1985	USDA-ARS	4b	A	A

Key	
Yellow	Nova Scotia, 1981
Green	Massachusetts, 1983
Blue	California, 1985
White	Control Strains

Letter designations represent a longer assigned string which uniquely identifies a strain "Pattern"

## RESULTS OVERVIEW

- The multi-locus tNGS genotyping method correctly identified the species of 25 *Listeria* isolates tested.
- For 24 of the *Listeria* isolates tested and typable by the system, a unique, identification number was assigned.
- Excellent concordance between outbreak collection metadata and genotyping data: Cohen's Kappa of 0.891.
- Moderately high discriminatory power of the methodology used: Hunter Gaston Discriminatory Index of 0.8369.

## CONCLUSIONS

- This study highlights the capability of an end-to-end NGS platform to quickly and accurately distinguish between *Listeria monocytogenes* subtypes as well as at the species level.
- "Patterns" assigned to distinct genotypes can be leveraged to efficiently track the movement of *Listeria* contamination within a facility.



## References

Ducey TF, Page B, Usgaard T, Borucki MK, Pupedis K, Ward TJ. A single-nucleotide-polymorphism-based multilocus genotyping assay for subtyping lineage I isolates of *Listeria monocytogenes*. *Appl Environ Microbiol*. 2007 Jan;73(1):133-47. doi: 10.1128/AEM.01453-06. Epub 2006 Nov 3. PMID: 17085705; PMCID: PMC1797101.

Wesley IV, Ashton F. Restriction enzyme analysis of *Listeria monocytogenes* strains associated with food-borne epidemics. *Appl Environ Microbiol*. 1991 Apr;57(4):969-75. doi: 10.1128/aem.57.4.969-975.1991. PMID: 1905523; PMCID: PMC182831.

